

# Determining the Frequency that Leads to Optimal Detection of Cell Membrane Permeabilization

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## Introduction

- In electroporation (EP), electric pulses are applied to cells to cause temporary cell membrane permeabilization, allowing delivery of molecules into cells such as DNA
- Cells must receive an appropriate pulse for successful, functional delivery
  - If the field is too weak or too short, the cell will not be permeabilized
  - If the field is too strong or too long, the cell will not recover and will die
  - The "correct" pulse will vary between cell types and even from cell-to-cell
- Our lab is currently developing a continuous flow microfluidic system that detects when individual cells become permeabilized to avoid over exposure to the electric field. This is to avoid killing the cell.
  - Monitor changes in the electrical impedance of cells by measuring the current across the microfluidic channel
- The goal of this study is to identify the optimal frequency at which to monitor different cell types to detect permeabilization with the greatest sensitivity

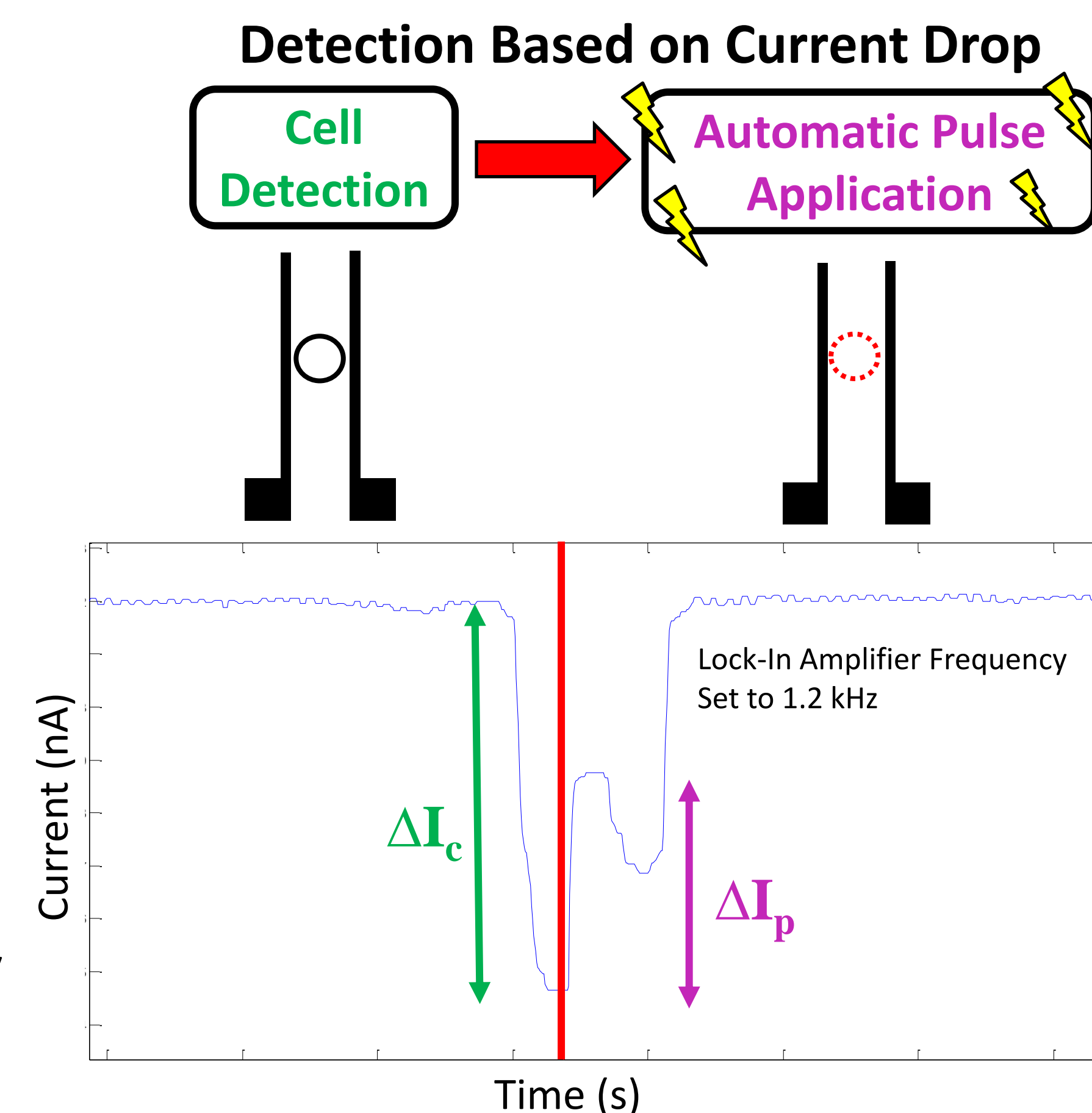
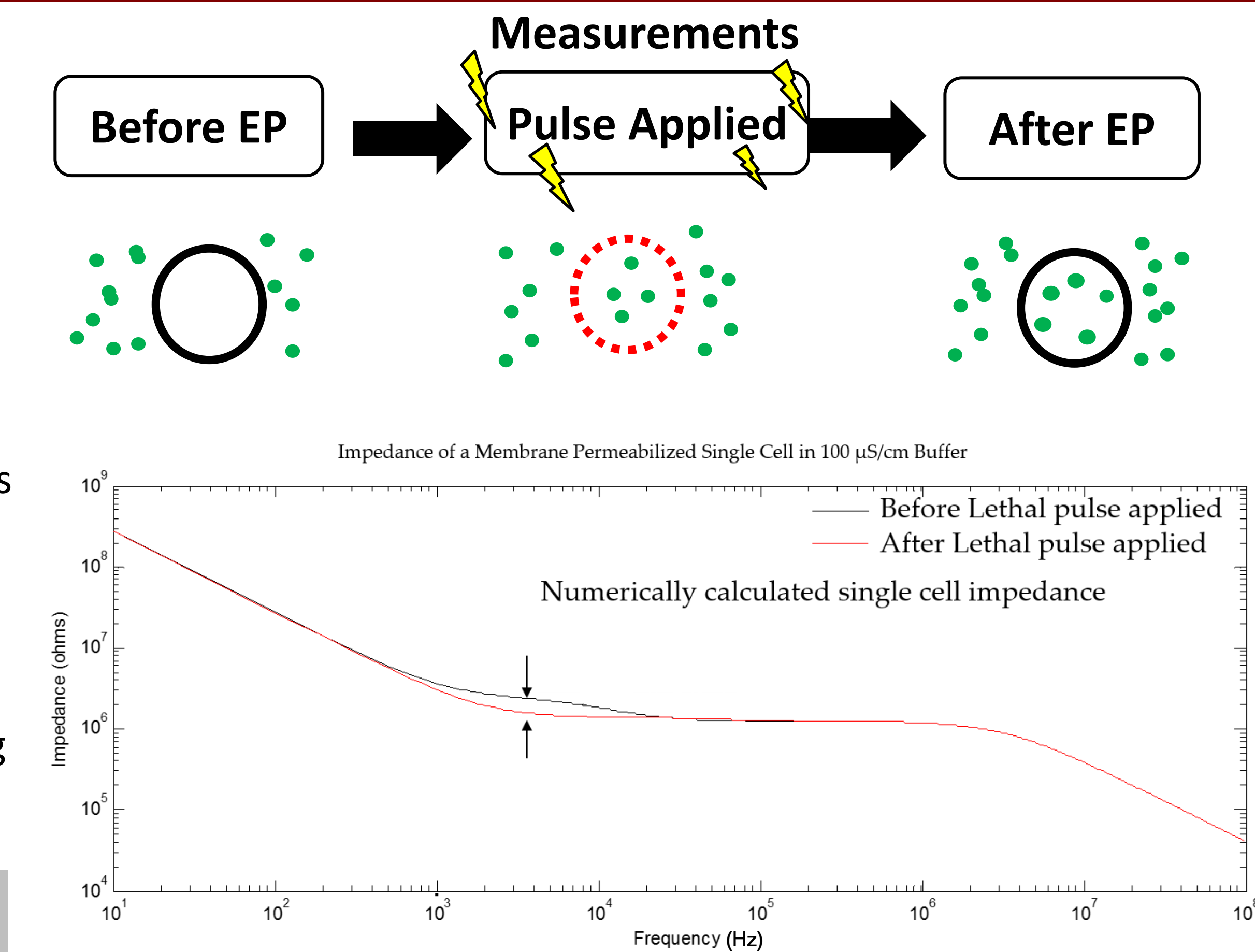


Figure 1: Current displacement due to cell entering region between electrodes and permeabilization

## Rationale

- Cells are suspended in a conductive buffer solution
- Electrodes measure the current across the microfluidic channel
- When a cell enters between the electrodes, the current drops ( $\Delta I_c$ ) because of the increase in impedance due to the presence of the cell
  - This drop triggers pulse application (red line)
- When the cell is permeabilized, the current increases ( $\Delta I_p$ ) because of the decrease in cell impedance
- We have modeled how this change in impedance depends on frequency and measure the change in NIH 3T3 fibroblast cells
  - There is an optimal frequency for detecting permeabilization

Research question: Does the optimal frequency to detect changes in impedance depend on the cell type?



## Experimental Approach

Prior to each experiment, cells are suspended in a conductive buffer solution and placed into a syringe. This syringe is loaded into a syringe pump and connected to the microdevice through thin tubing. The microdevice is placed onto a microscope. The syringe pump is used to control the flow rate of the cells in the microdevice.

### 1) Before Electroporation:

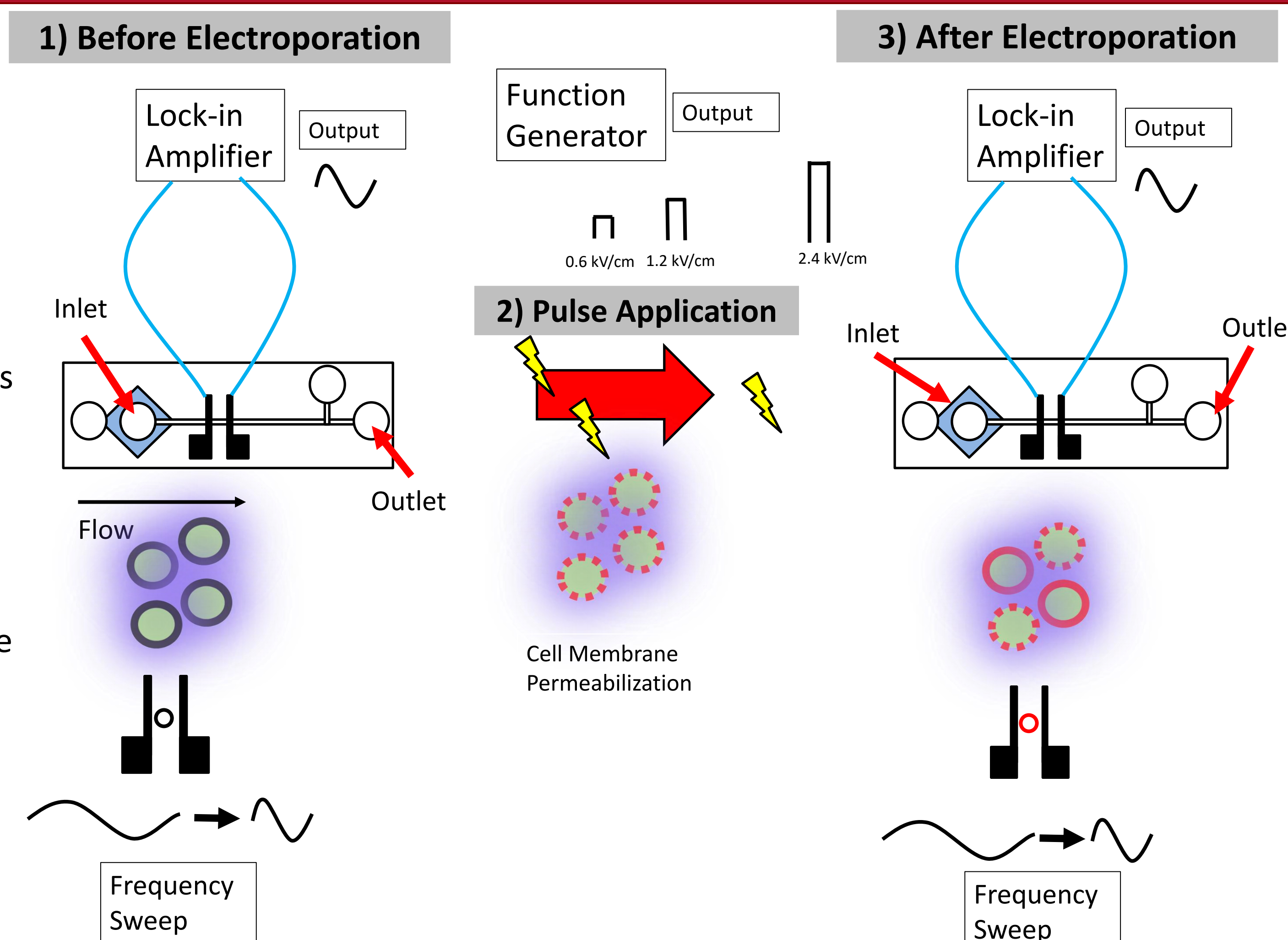
- Frequency sweep performed once a single cell is trapped between the electrodes
- Lock-in Amplifier measures the impedance at a set range of frequencies

### 2) Pulse Application:

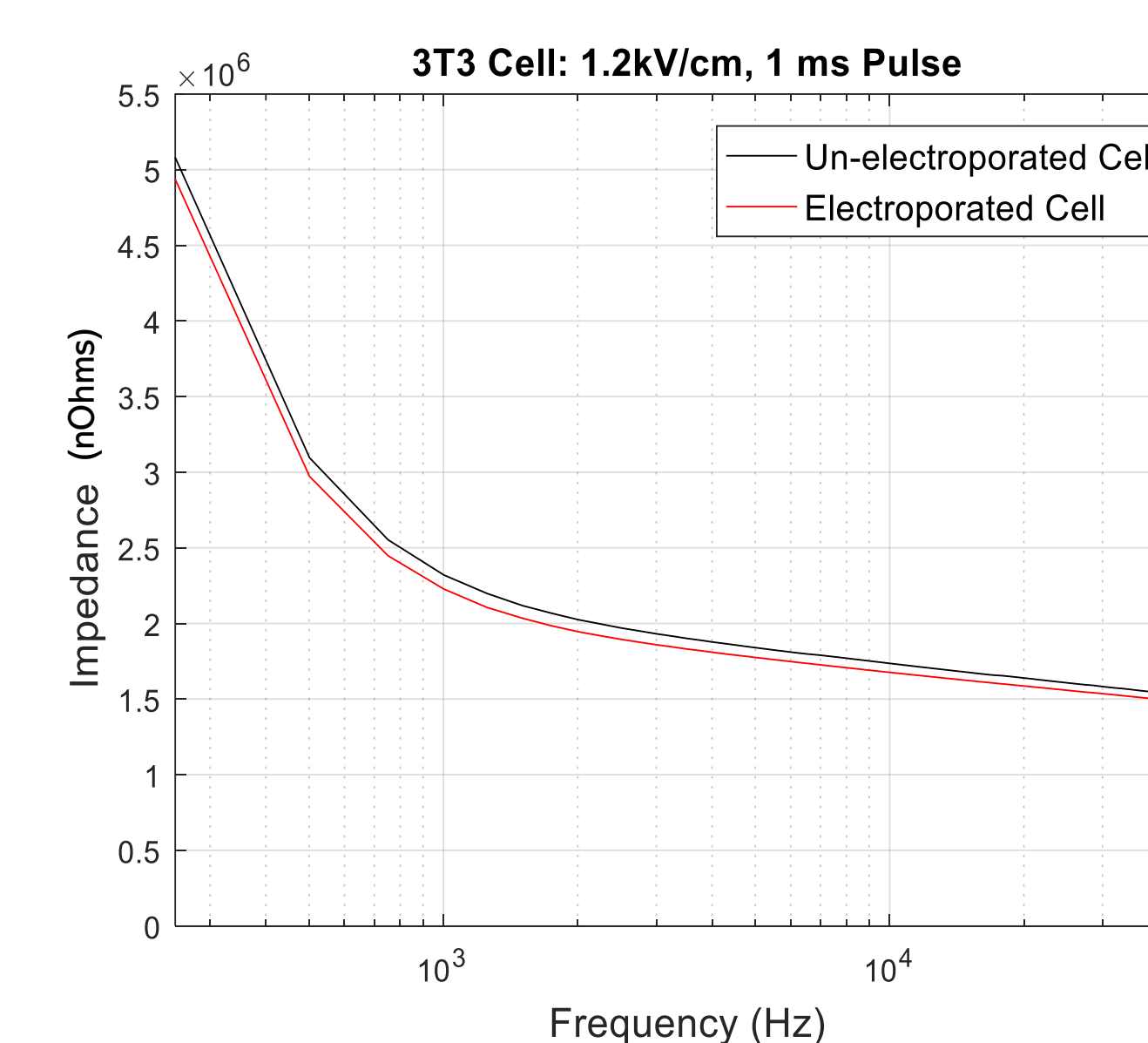
- Function generator outputs a square wave pulse
- Pulse strength used was 0.6 kV/cm, 1.2 kV/cm, and 2.4 kV/cm depending on the experiment
- Pulse duration is kept constant at 1 ms

### 3) After Electroporation:

- Lock-in Amplifier performs second frequency sweep

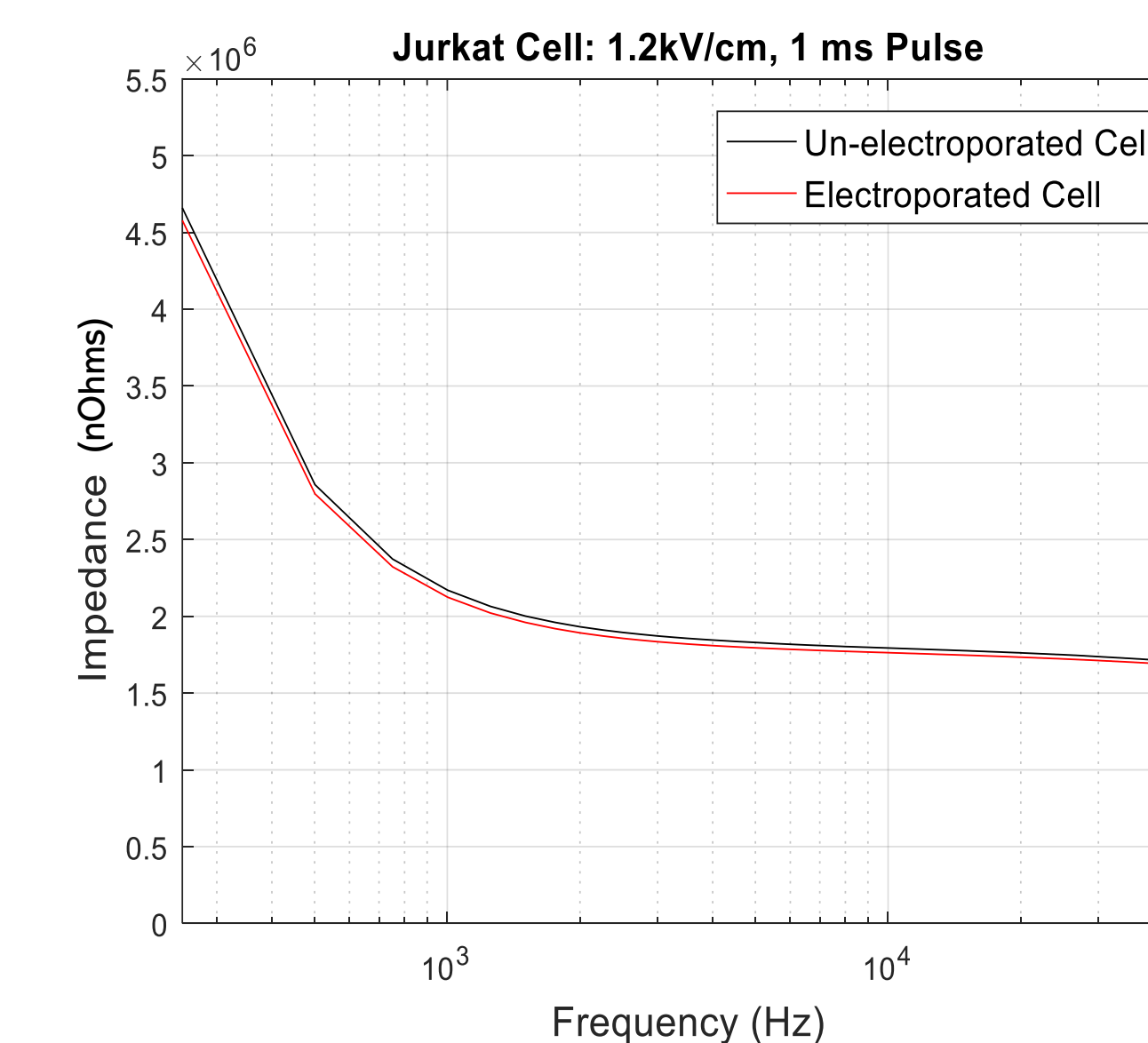


## Results



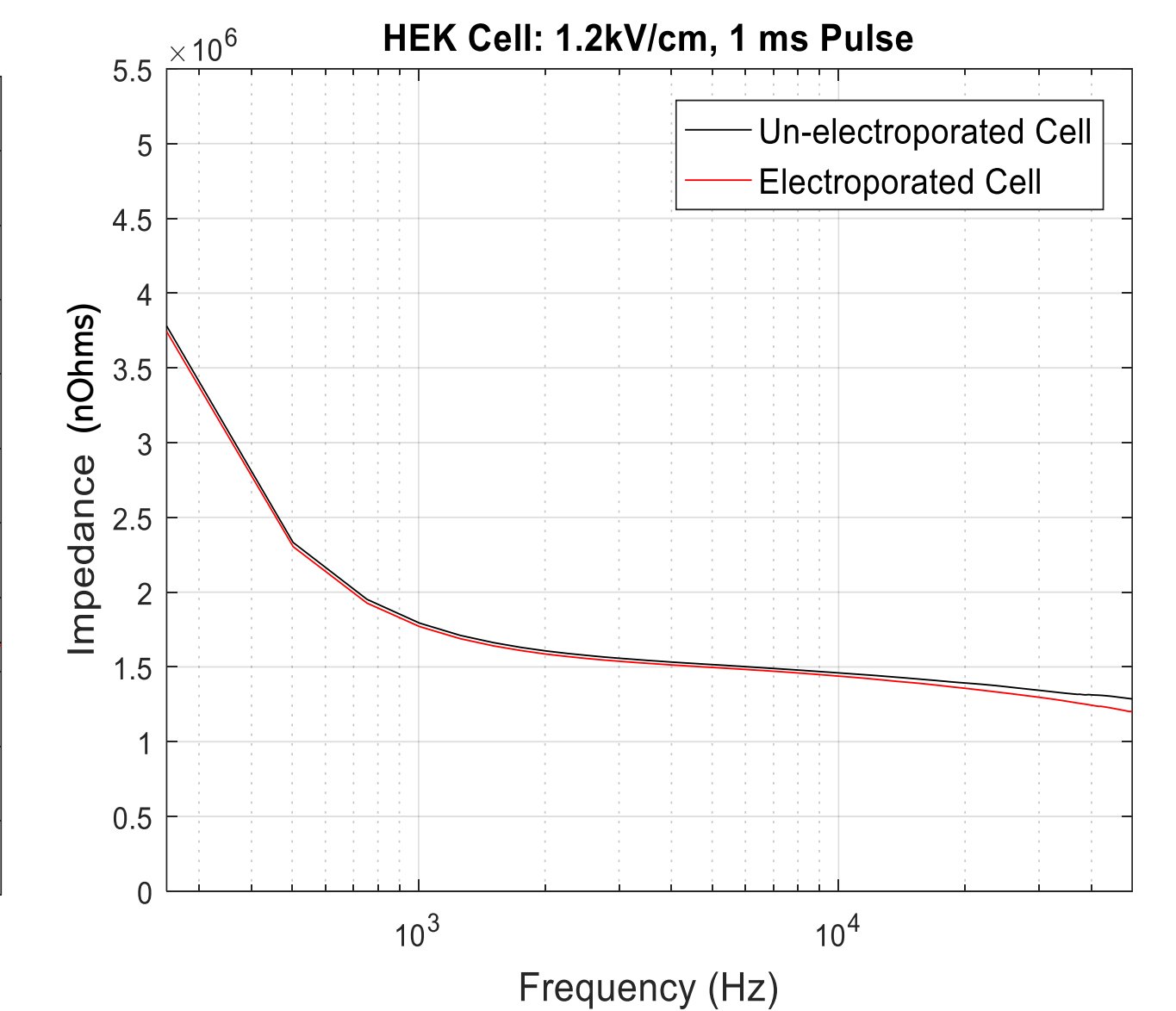
### 3T3 cells:

- Maximum percent change within frequency range 1->10 kHz: **4.21%**
- Corresponding frequency: **1250 Hz**



### Jurkat cells:

- Maximum percent change within frequency range 1->10 kHz: **2.10%**
- Corresponding frequency: **1000 Hz**



### HEK293 cells:

- Maximum percent change within frequency range 1->10 kHz: **1.49%**
- Corresponding frequency: **1000 Hz**

## Discussion

- Data on these three cell types suggests that optimal frequency for detection is around 1 kHz
- Difference in percent change between different cell types is possibly due to differences in cell size

## Future Direction

- Increase the sample size on each cell type
- Collect impedance data on additional pulsing conditions
- Expand the variety of cell types

## Acknowledgements

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## References

Zheng, M., Sherba, J. J., Shan, J. W., Lin, H., Shreiber, D. I., & Zahn, J. D. (2017). Continuous-flow, electrically-triggered, single cell-level electroporation. *Technology*, 05(01), 31-41. doi:10.1142/S2339547817500017