- The instrument can analyze particle size ranges from 0.4 µm to 40 µm diameter.
- Maximum Acquisition Rate: 25,000 events per second.

Gallios Laser Specifications:

Lasers	Power output
Blue Solid State Diode, 488nm	22 mW
Red Solid State Diode, 638nm	25 mW
Violet Solid State Diode, 405nm	40 mW

Gallios Configuration:

Excitation	Detectors	Emission	Fluorochromes
488nm	FL1	525/20nm	FITC, GFP, YFP, AF488, CFSE, DCFH
	FL2	575/20nm	PE, DsRed, Vybrant orange
	FL3	620/30nm	PI, PE-Texas Red, PECF594, PE-Vio615
	FL4	675/20nm	PE-Cy5, PerCP, 7AAD
		OR 695/30nm	PECy5.5, PerCPeF710, PerCPVio700
	FL5	755LP nm	PE-Cy7, PEVio770
633nm	FL6	660/20nm	APC, AF647, eF660
	FL7	725/20nm	AF700
	FL8	755LP nm	APC-Cy7, APC-Alexa750, APC-Alexa770, APC-eF780
405nm	FL9	450/50nm	BV421, Pac Blue, AF405, VioBlue, DAPI
	FL10	550/40nm	Krome Orange, Pacific Orange, AF430, VioGreen

Available software for analyzing data:

- Kaluza Analysis Software. You can analyze your data on your computer using Kaluza analysis software. First download the one-month free demo of Kaluza software here. Second, ask us to borrow a dongle for your computer to use the Kaluza analysis software. To download Kaluza Instructions click here.
- ModFit LT, Verity Software House, Inc.

Tips for preparation of samples:

- Cell concentration in each tube: about 1 million cells/ml
- Sample volume in each tube: 0.3 – 1 ml (minimum volume is 0.3 ml)
- Sample tubes must be polypropylene (12×75 mm round bottom polypropylene tube- BD Falcon Cat# 352002 for non sterile, or Cat# 352063 for sterile).
- All samples should be filtered with 40 µm nylon cell strainers (Falcon # 352340) prior to running to avoid clogging the machines.
- If you have samples stained with more than one fluorochrome per sample, we recommend you bring single color stained positive control samples (each fluorochrome individually) and unstained or negative control (isotype) cells for proper compensation.
- We recommend that samples are kept on ice or chilled until ready to run on the instrument.